

Proceedings of the Zoological Institute RAS Vol. 315, No. 3, 2011, pp. 352–364

УДК 597

MORPHOLOGICAL AND mtDNA SEQUENCE STUDIES SEARCHING FOR THE ROOTS OF SILVER CRUCIAN CARP *CARASSIUS GIBELIO* (CYPRINIDAE) FROM PONDS OF SERGIEVKA PARK, SAINT PETERSBURG, RUSSIA

H. Sakai^{1*}, Y. Yamazaki², M.V. Nazarkin³, V.G. Sideleva³, D.A. Chmilevsky⁴, K. Iguchi⁵ and A. Goto⁶

¹National Fisheries University, Nagata-honmachi 2-7-1, Shimonoseki, Yamaguchi 759-6595, Japan; e-mail: sakaih@fish-u.ac.jp

²University of Toyama, 3190 Gofuku, Toyama 930-8555, Japan; e-mail: yatsume@sci.u-toyama.ac.jp

³Zoological Institute of the Russian Academy of Sciences, Universitetskaya Emb. 1, 199034 Saint Petersburg, Russia; e-mail: vsideleva@gmail.com

⁴Biological Research Institute, Saint Petersburg State University, Oranienbaumskoe Shosse 2, 198510 Petergof, Russia; e-mail: ichtyol1943@mail.ru

⁵National Research Institute of Fisheries Science, Komaki 1088, Ueda, Nagano 386-0031, Japan, e-mail: keyichi@affrc.go.jp ⁶Field Science Center for Northern Biosphere, Hokkaido University, Minatocho 3-1-1, Hakodate, Hokkaido 041-8611, Japan; e-mail: akir@fish.hokudai.ac.jp

ABSTRACT

The roots of the silver crucian carp *Carassius gibelio* from ponds of Sergievka Park (Petergof, Saint Petersburg), were surveyed based on morphological and molecular (mitochondrial DNA sequences, control and cytochrome *b* regions) study of samples of this species, collected there in 1930-thies and 2008, in comparison with *C. g. gibelio* and *C. gibelio*. subsp. M. collected from Kazakhstan. It is demonstrated, that the fish is most similar to *C. gibelio gibelio* morphologically. Their mitochondrial haplotypes were clustered with those of the Far Eastern populations such as the Amurian one. The result indicates that the silver crucian carp of Sergievka Park has been introduced originally from the Far East region.

Key words: crucian carp, morphology, mtDNA, Prussian carp

ИЗУЧЕНИЕ МОРФОЛОГИИ И ПОСЛЕДОВАТЕЛЬНОСТИ MTДНК СЕРЕБРЯНОГО КАРАСЯ *CARASSIUS GIBELIO* (CYPRINIDAE) ИЗ ПРУДОВ ПАРКА СЕРГИЕВКА (САНКТ-ПЕТЕРБУРГ, РОССИЯ) С ЦЕЛЬЮ ВЫЯСНЕНИЯ ЕГО ПРОИСХОЖДЕНИЯ

Х. Сакаи^{1*}, И. Ямазаки², М.В. Назаркин³, В.Г. Сиделева³, Д.А. Чмилевский⁴, К. Игучи⁵, А. Гото⁶

¹National Fisheries University, Nagata-honmachi 2-7-1, Shimonoseki, Yamaguchi 759-6595, Japan; e-mail: sakaih@fish-u.ac.jp

²University of Toyama, 3190 Gofuku, Toyama 930-8555, Japan; e-mail: yatsume@sci.u-toyama.ac.jp

³Зоологический институт Российской академии наук, Университетская наб. 1, 199034 Санкт-Петербург, Россия; e-mail: vsideleva@gmail.com

⁴Биологический научно-исследовательский институт, Санкт-Петербургский государственный университет, Ораниенбаумское ш. 2, 198510 Петергоф; e-mail: ichtyol1943@mail.ru

⁵National Research Institute of Fisheries Science, Komaki 1088, Ueda, Nagano 386-0031, Japan, e-mail: keyichi@affrc.go.jp ⁶Field Science Center for Northern Biosphere, Hokkaido University, Minatocho 3-1-1, Hakodate, Hokkaido 041-8611, Japan; e-mail: akir@fish.hokudai.ac.jp

*Corresponding author / Автор-корреспондент

РЕЗЮМЕ

Изучено происхождение *Carassius gibelio*, интродуцированного в пруды парка Сергиевка (Петергоф, Санкт-Петербург) на основе сравнительно-морфологического и молекулярно-генетического (участки контрольного региона и цитохрома *b* мтДНК) изучения выборок серебряного карася, собранных в этих прудах в 1930-х и 2008 гг., в сравнении с *C. gibelio gibelio* и *C. gibelio* subsp. М из водоемов вблизи г. Семипалатинска (Казахстан). Показано, что по морфологическим признакам рыбы из прудов парка Сергиевка наиболее сходны с *C. gibelio gibelio*. Полученные гаплотипы *C. gibelio* сходны с таковыми карасей из реки Амур и других рек Дальнего Востока. Следовательно, серебряный карась вселен в пруды парка Сергиевка из водоемов Дальневосточного региона России.

Ключевые слова: серебряный карась, морфология, мтДНК, прусский карась

INTRODUCTION

Crucian carp, the genus *Carassius* Nilsson, 1832 is well known for its controversial taxonomy and contentious morphological classification (Berg 1949; Nakamura 1969). At least three major species have been recognized: common crucian carp C. carassius (Linnaeus, 1758), mainly from Europe; silver crucian carp C. auratus (Linnaeus, 1758), mainly from Asia; and Japanese crucian carp C. cuvieri Temminck et Schlegel, 1846 from Lake Biwa, Japan (Berg 1949; Nakamura 1969; Hosoya 2002). The main taxonomic controversy has centered on *C. auratus*, because of its variation in morphology as well as in ploidy size (Nakamura 1969). In the Eurasian Continent, however, two sub-species have been recognized traditionally: Prussian carp C. auratus gibelio (Bloch, 1782) from Germany and Poland to the Amur River system, and C. auratus auratus from China (Berg 1949; Wu 1977), with some authors treating them as the same subspecies, C. auratus gibelio and some as distinct species (Hensel 1971; Kottelat and Freyhof 2007; Kalous et al. 2007).

Because the species name "auratus" (Linnaeus 1758) was established based on the domestic goldfish with a tri-lobed tail, International Commission on Zoological Nomenclature (2003) has recommended that the name should be limitedly applied to the goldfish. Sakai et al. (2009) have discussed that the name "gibelio" proposed by Bloch (1782) from Prussia should be used for the wild form instead of "auratus". However, it has been claimed that ongoing introductions of Far Eastern crucian carp to Europe have brought about considerable propagation of the Far Eastern form in European waters (Hensel 1971; Holčík and Žitňan 1977; Cherfas 1979), resulting in much difficulty in proving the existence of native European Prussian carp (Hensel 1971). Moreover,

the type specimen of *Cyprinus gibelio* Bloch (1782) has been lost (Paepke 1999) and it is now difficult to judge which type of morphology should be identified as the original "*gibelio*", the question having been unsolved yet.

By the way, Sakai et al. (2009) have found a new stock of silver crucian carp from Kazakhstan, tentatively called *C. gibelio* subsp. M, with a peculiar mitochondrial haplotype different from those ever known and with an intermediate number of gillrakers between *C. auratus* or *gibelio* and *C. carassius*. They suggested its wider distribution in Central Eurasia such as from the Lena River system (Kirillov 1956) to Saint Petersburg (Grib 1935) based on the morphological characteristics described. They also discussed a possibility that *C. gibelio* subsp. M may represent the true Prussian carp *C. gibelio* described by Bloch (1782), if the fish from European Russia appears to be the same stock with *C. gibelio* subsp. M.

Recently, Takada et al. (2010) conducted a comprehensive mitochondrial gene sequence analysis of C. auratus or gibelio. They revealed the existence of two mitochondrial superlineages; one including three lineages from Japan (clades I-III) and the other including four, one from the Ryukyu Islands (clade IV), another from Taiwan Island (clade VI) and the other two from Eurasian Continent (clades V and VII). Among them, one of the Eurasian lineages (clade V) contained a monophyletic cluster of three cytochrome b sequences recorded from the Czech Republic. Takada et al. (2010) considered them to be native to Europe, although all the other fish registered from outside East Asia were judged as introduced from there. However, they did not touch C. gibelio subsp. M sensu Sakai et al. (2009), and then it is still unknown whether or not the same stock as C. gibelio subsp. M is distributed in other regions than Kazakhstan, especially in Europe.

The authors collected crucian carp from ponds of Sergievka Park (Petergof, Saint Petersburg), where Grib (1935) reported the fish that is morphologically similar to C. gibelio subsp. M sensu Sakai et al. (2009). Subsequent studies aim to determine the roots of the fish from Sergievka Park based on morphological characters and mitochondrial DNA (mtDNA) sequence analyses, anticipating a discovery of the true Prussian carp. The consequence of the research is that the fish seem not to be native, but to have been introduced stock possibly from the Far East region.

MATERIAL AND METHODS

Fish collection. Silver crucian carp were collected from ponds of Sergievka Park initially settled in 18th century (Vlasov 2005) in 2008 (Fig. 1). The fish were initially identified following the keys given by Berg (1949), being 24 individuals of C. gibelio (Fig. 2A). Three individuals of common crucian carp C. carassius were also collected but released soon after. Immediately after collection, nine fish were anesthetized and a drop of blood from the first gill arch was placed on a slide for measuring erythrocyte size. A piece of pectoral fin was clipped and fixed



Fig. 1. Map showing the location of Sergievka Park, Saint Petersburg, Russia.

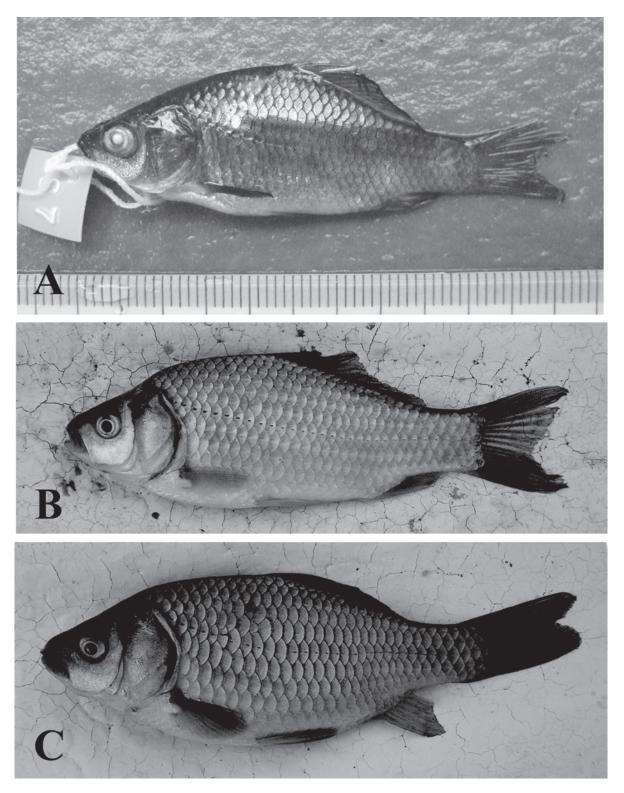


Fig. 2. Specimens of *Carassius gibelio*: (A) *C. gibelio* collected from Sergievka Park, 44.9 mm standard length (SL); (B) *C. g. gibelio* from Kazakhstan, 95.5 mm SL; (C) *C. g.* subsp. M from Kazakhstan, 98.8 mm SL.

H. Sakai et al.



Fig. 3. Three specimens of Carassius gibelio, 110.8–125.6 mm SL, collected from Sergievka Park by Grib (1935).

356

Table 1. Samples of silver crucian carps genus *Carassius* from Sergievka Park (Russia), and Kazakhstan. Designations: * – significantdifference from fish collected from Sergievka Park in the present study (p < 0.05, Student's t-test); ZIN – catalogue number registered inZoological Institute of the Russian Academy of Sciences; HUMZ – catalogue number registered in Hokkaido University, Laboratory ofMarine Zoology, Faculty of Fisheries. For other designations see Material and Methods.

| | Sergievka Park | | Kazakhstan (Sakai et al. 2009) | |
|----------|----------------|--------------------------|--------------------------------|---------------|
| No. ind. | Present study | Grib (1935) | C. gibelio gibelio | C.g. subsp. M |
| | 24 | 3 | 19 | 20 |
| SL(mm) | 31.3-52.4 | 110.8-125.6 | 66.1-127.9 | 42.9-104.5 |
| | In % of SL (m | ean with standard deviat | ion in parenthesis) | |
| BD | 41.1 (3.2) | 47.8 (1.7) | 30.2 (2.3) | 40.3 (1.7) |
| HL | 33.2 (1.3) | 31.3 (2.5) | 30.4 (1.4) | 32.1 (1.1) |
| CPD | 15.2 (0.7) | 16.0 (0.3) | 15.1 (0.8) | 14.1 (0.8) |
| PreDL | 54.0 (2.2) | _ | 52.8 (1.8) | 54.6 (2.2) |
| PrePL | 52.7 (1.9) | - | 50.9 (1.6) | 53.2 (1.5) |
| PreAL | 78.2 (2.0) | - | 77.3 (2.6) | 78.0 (1.9) |
| | | In % of HL | | |
| SnL | 29.4 (2.2) | 30.6 (2.5) | 24.6 (1.6) | 24.7 (1.5) |
| ED | 28.4 (6.3) | 17.8 (1.3) | 21.0 (2.0) | 21.4 (2.0) |
| UJL | 23.8 (2.2) | - | 21.6 (1.6) | 24.9 (1.7) |
| V | 29.4 (0.6) | - | 30.4 (0.5)* | 30.9 (0.6)* |
| D | 16.4 (0.7) | 15.0 (0.0)* | 17.0 (0.5)* | 15.6 (0.7)* |
| А | 5.0 (0.0) | 5.0 (0.0) | 5.0 (0.0) | 5.0 (0.0) |
| Ll | 28.0 (0.8) | 32.5 (0.7)* | 30.3 (0.5)* | 30.6 (0.8)* |
| Sal | 6.1 (0.6) | 6.0 (0.0) | 5.9 (0.4) | 5.0 (0.0)* |
| Sbl | 5.2 (0.4) | 5.0 (5.0) | 5.3 (0.5) | 4.9 (0.3)* |
| Gr | 36.9 (1.5) | 45.0 (2.8)* | 46.7 (1.1)* | 38.5 (2.1)* |
| ZIN | - | 25220 | 54385 | 54384 |
| HUMZ | 209617-209640 | _ | 209587-209601 | 209572-209586 |

in 99% ethanol for DNA extraction. The remaining bodies and the rest of the fish were labeled and fixed in 70% ethanol for morphological observation.

Morphological analyses. Meristics, known to be variable among both species and sub-species (Na-kamura 1969), were selected as follows: vertebrae, inclusive of Weberian apparatus (V), branched dorsal fin rays (D), branched anal fin rays (A), scales on lateral line series (Ll), scales on transverse series from dorsal fin base to above lateral line (*Sal*), scales on transverse series from anal fin base to below lateral line (*Sbl*), and gill rakers on first gill arch (*Gr*). Morphometrics were measured as follows: standard length (*SL*), body depth (*BD*), head length (*HL*), caudal peduncle depth (*CPD*), pre-dorsal length (*PreDL*),

pre-anal length (*PreAL*), snout length (*SnL*), eye diameter (*ED*), and upper jaw length (*UJL*).

The fish from the same locality collected by Grib (1935; Fig. 3) and *C. gibelio gibelio* (Fig. 2B) and *C. gibelio* subsp. M (Fig. 2C) from Kazakhstan collected by Sakai et al. (2009) were morphologically compared with the present fish, their catalogue numbers being indicated in Table 1. Difference from the fish of Sergievka Park in meristics, which are possibly hardly affected by relative growth, was tested by Student's t-test.

Erythrocyte diameter of ten cells (long and short axes) of each fish was measured utilizing a micrometer attached to a microscope, and mean μ m² was calculated, such possibly being an indicator of relative

Table 2. Samples of crucian and common carps used for mtDNA analysis. Designation: * – accession number in DDBJ/GenBank/EMBLDNA Data Bank.

| Access. N.* | Locality | Literature | |
|-------------|--------------------------------------------------|--------------------------------|--|
| | Control region | | |
| | Carassius gibelio from Saint Petersbu | urg | |
| AB587672 | Sergievka Park, Saint Petersburg, Russ | ia Present study | |
| | C. gibelio subsp. | | |
| AB274413 | Mikhailovski Lake, Kazakhstan | Sakai et al. (2009) | |
| | C. gibelio gibelio | | |
| AB274414 | Chara River, Kazakhstan | Sakai et al. (2009) | |
| AB274415 | Chara River, Kazakhstan | Sakai et al. (2009) | |
| AB274416 | Chara River, Kazakhstan | Sakai et al. (2009) | |
| AB080012 | Amur River, Russia | Yamamoto et al. (2010) | |
| AB080013 | Amur River, Russia | Yamamoto et al. (2010) | |
| AB080014 | Amur River, Russia | Yamamoto et al. (2010) | |
| AY940117 | Labe River, Czech Republic | Mendel et al. (unpubl.) | |
| AY940118 | Marava River, Czech Republic | Vetesnik et al. (2007) | |
| AY940119 | Marava River, Czech Republic | Mendel et al. (unpubl.) | |
| AJ388413 | France | Gilles and Lecointre (unpubl.) | |
| | C. gibelio langsdorfii | | |
| AB079925 | Kasumigaura Lake, Japan | Iguchi et al. (2003) | |
| AB079905 | Biwa Lake, Japan | Yamamoto et al. (2010) | |
| | C. gibelio grandoculis | | |
| AB079901 | Biwa Lake, Japan | Yamamoto et al. (2010) | |
| AB079903 | Biwa Lake, Japan | Yamamoto et al. (2010) | |
| | C. gibelio buergeri | | |
| AB079971 | Kako River, Japan | Yamamoto et al. (2010) | |
| AB079965 | Chikugo River, Japan | Yamamoto et al. (2010) | |
| | C. gibelio subsp. 1 | | |
| AB079951 | Urano River, Japan | Yamamoto et al. (2010) | |
| AB079940 | Mikata Lake, Japan | Yamamoto et al. (2010) | |
| 10013040 | | | |
| | C. gibelio subsp. 2 | | |
| AB079921 | Sakasazawa River, Japan | Iguchi et al. (2003) | |
| AB079929 | Kasumigaura Lake, Japan | Iguchi et al. (2003) | |
| | C. gibelio subsp. A (traditionally named C. auro | atus auratus) | |
| AB080009 | Yangtze River, China | Yamamoto et al. (2010) | |
| AB080010 | Yangtze River, China | Yamamoto et al. (2010) | |
| AB080011 | Yangtze River, China | Yamamoto et al. (2010) | |
| | C. cuvieri | | |
| AB080015 | Biwa Lake, Japan | Iguchi et al. (2003) | |
| | C. auratus (Goldfish) | | |
| AB052332 | Fish dealer | Murakami et al. (2001) | |

358

Access. N.* Locality Literature C. carassius AB274417 Mikhailovski Lake, Kazakhstan Sakai et al. (2009) DQ007649 Marava River, Czech Republic Mendel et al. (unpubl.) Cyprinus carpio (Common Carp) X61010 Taiwan Chang et al. (1994) Cytochrome b Carassius gibelio from Saint Petersburg AB605597 Sergievka Park, Saint Petersburg, Russia Present study C. gibelio gibelio AB368700 Amur River, Russia Takada et al. (2010) AB368701 Amur River, Russia Takada et al. (2010) AB368707 Ryukyu Islands Takada et al. (2010) GU170401 Amur River, China Liang et al. (unpubl.) DQ399926 Elbe River, Czech Republic Kalous et al. (2007) DQ399935 Elbe River, Czech Republic Kalous et al. (2007) DQ399940 Elbe River, Czech Republic Kalous et al. (2007)

ploidy size. The data of *C. carassius* (diploidy) and *C. gibelio* subsp. M (triploidy) from Kazakhstan (Sakai et al. 2009) were used for comparative purpose.

Sex was determined by a visual examination of the gonads.

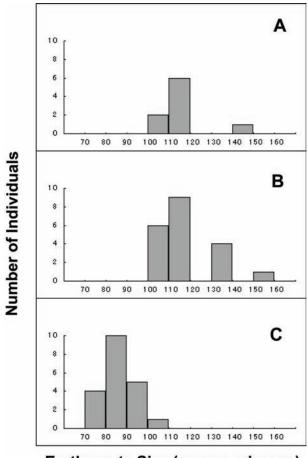
Mitochondrial analyses. Total genomic DNA was extracted from the fin samples, which were digested by proteinase K and purified by standard phenol/ chloroform extraction. The first one-third of the control region and the cytochrome b region of mtDNA were amplified with primers devised by Iguchi et al. (2003) and Takada et al. (2010), respectively. PCR amplification was conducted according to standard protocols with GeneAmp PCR 9700 (Applied Biosystems Inc., Foster City, CA, USA). Amplified double-stranded DNA was sequenced on an automated DNA sequencer ABI 310 (Applied Biosystems Inc., Foster City, CA, USA) with the amplification primers using the BigDye Terminator cycle sequencing kit ver. 3.1 (Applied Biosystems Inc., Foster City, CA, USA). Registered accession numbers in the DDBL/GenBank/EMBL DNA Data Bank were shown in Table 2.

To determine the relationships between the present crucian carp and those from other regions, a phylogenetic analysis was conducted incorporating data already registered with the DDBJ/GenBank/ EMBL DNA Data Bank, most of which are the same data already been analyzed by Sakai et al. (2009) and Takada et al. (2010) (Table 2). Alignment of multiple sequences was done using Clustal W 1.6 (Thompson et al. 1994). A neighbor-joining tree (Saitou and Nei 1987) with 1000 bootstrap replications (Efron 1979) was constructed using PAUP* 4.0b10 (Swofford 2002), based on 283 bp excluding insertions/ deletions for the control region and 1141 bp for the cytochrome b region, with the model of Hasegawa et al. (1985) selected by ModelTest ver. 3.7 (Posada and Crandall 1998).

RESULTS

Morphological differences. Morphological comparison is shown in Table 1. The fish from Sergievka Park were the smallest of the samples, and relative measurements such as BD, HL, ED, etc. were differ-

Table 2. Continued.



Erythrocyte Size (square microns)

Fig. 4. Erythrocyte size (square microns) of *Carassius gibelio* from Sergievka Park (A), *C. g. gibelio* (B) and *C. carassius* (C) from Kazakhstan.

ent even from those of the same locality collected by Grib (1935).

Meristics of the fish from Sergievka Park, except for anal fin rays (A), were significantly different from C. gibelio subsp. M from Kazakhstan. Dorsal fin rays (D), gill rakers (Gr) and lateral line scales (Ll) were also different from those of all the other samples. On the other hand, transverse scales above (Sal) and below the lateral line (Sbl) were not different from the fish by Grib (1935) as well as C. gibelio gibelio from Kazakhstan.

Erythrocyte size and sex. The mean erythrocyte sizes were more than 100 μ m² in the fish from Sergievka Park and *C. gibelio* subsp. M from Kazakhstan, and less than 110 μ m² in *C. carassius* from Kazakhstan (Fig. 4A, 4B, 4C, respectively).

H. Sakai et al.

All nine individuals of fish from Sergievka Park were too young to determine their sex by eyes, whereas all 20 individuals of *C. gibelio* subsp. M from Kazakhstan were females and six out of 20 *C. carassius* from Kazakhstan were males.

Phylogenetic analysis. The haplotypes of *C. carassius* formed a species cluster based on control region sequences (Fig. 5). The remaining haplotypes were included in the *C. gibelio* group (bootstrap probability 76%), being divided into three; *C. gibelio* subsp. M from Kazakhstan, CHINA-AMUR (87%) and JAPAN clusters (less than 50%). The CHINA-AMUR cluster was subdivided into two clusters, one mainly from China (96%, clade VII by Takada et al. 2010) and the other mainly from Amur (99%, clade V by Takada et al. 2010), the fish from Sergievka Park being included in the latter.

According to the result based on cytochrome b, the sequence of fish from Sergievka Park was just the same as that of one haplotype from Czech Republic (Fig. 6). Moreover, the sequence registered from the Chinese Amur was also the same as them. The three sequences recorded from Czech Republic, consequently, did not form their own cluster anymore.

DISCUSSION

The silver crucian carp from ponds of Sergievka Park was morphologically more similar to *Carassius gibelio gibelio* than *C. gibelio* subsp. M sensu Sakai et al. (2009) from Kazakhstan, whereas the number of gill rakers was smaller than the former and near to the latter. The number of gill rakers in cyprinids is known to increase according to growth (Nakamura 1969). The smaller number of gill rakers in the present sample, therefore, is possibly due to their smaller body size.

Erythrocyte size shows that the fish from Sergievka Park are possibly triploid (or more) as *C. gibelio* subsp. M from Kazakhstan in comparison with that of diploid *C. carassius*. Such triploid populations are rather common wherever silver crucian carp exists (Cherfas 1979).

Phylogenetic analysis indicates that the fish from Sergievka Park is not native but is possibly introduced stock originated from the Far East region. In fact, the introductions of the Far Eastern crucian carp to Europe via Russian waters have brought about considerable propagation of them in there (Hensel 1971; Holčík and Žitňan 1977; Cherfas 1979). The

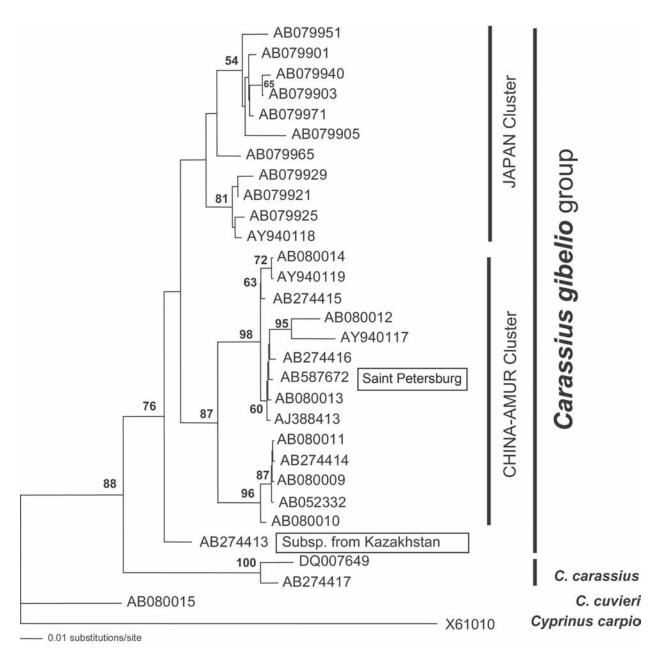


Fig. 5. Mitochondrial DNA phylogeny of crucian carps based on control region sequences by the neighbour-joining method with % bootstrap probabilities (1000 replications). See Table 2 for OUT names. Enclosed haplotypes are those of *Carassius gibelio* from Sergievka Park, Saint Petersburg, and *C. g.* subsp. M from Kazakhstan.

fish of Sergievka Park must have been one of such. According to Takada et al. (2010), two lineages exist in the Eurasian Continent, clade V and VII. One of them, clade V contained a monophyletic cluster of cytochrome b sequences from the Czech Republic, being considered to be native to Europe (Takada et al. 2010). The fish from Sergievka Park of the present study, however, showed just the same cytochrome b sequence as that of one haplotype of the Czech Republic cluster as well as that registered from the Amur River system. The cluster recorded from Czech Republic, consequently, is not unique to Europe

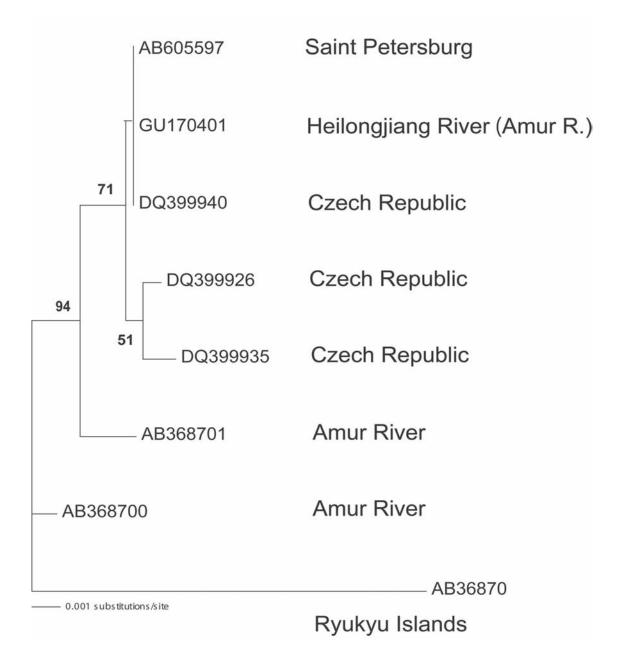


Fig. 6. Mitochondrial DNA phylogeny based on cytochrome *b* sequences by the neighbour-joining method with % bootstrap probabilities (1000 replications). See Table 2 for OUT names. Enclosed haplotype is that of *Carassius gibelio* from Sergievka Park, Saint-Petersburg. The haplotype from Ryukyu Islands (AB36870) is a member of clade VII, possibly being non-native to Ryukyu's, and the other haplotypes are members of clade V sensu Takada et al. (2010).

anymore. It would support the Amurian origin of the Czech fish as well as the Sergievka fish.

According to Bychowsky (1933), crucian carp had possibly been introduced to the ponds of Sergievka Park around 1913, but he did not describe from where. Grib (1935) himself also suggested the possibility that the fish had been introduced from Asia. Chmilevsky and Ivanov (2005) listed eight fish species inhabiting the ponds of Sergievka Park, including *Percottus glehni* that is also a member of the Amurian fish fauna (Berg 1949). Such introductions of alien species to the ponds may have been done several times.

Sakai et al. (2009) reported a new crucian carp stock *C. gibelio* subsp. M, seemingly native to Kazakhstan. Under the ambiguity of what the Bloch's (1782) "gibelio" is, they also discussed a possibility that the Kazakhstan fish may represent the true Prussian carp *C. gibelio* by Bloch (1782), if the fish from European Russia such as from Sergievka Park documented by Grib (1935) appears to be the same stock with *C. gibelio* subsp. M. However, morphological and genetic data clearly reject that possibility. The silver crucian carp from Sergievka Park is thought to have been an introduced stock originated from the Far East region such as the Amur River. It is still unknown, therefore, whether *C. gibelio* native to Europe exists or not.

ACKNOWLEDGMENTS

The authors thank the following persons for assistance: Y. Suda, K. Sakai, M. Sano, N. Shimogama, N. Yamamoto and M. Matsumoto. This work was supported by Grant-in-Aid for Scientific Research (B) (19405011), Japan Society for the Promotion of Science. This work was supported by Ministry of Education and Science of the Russian Federation.

REFERENCES

- Berg L.S. 1949. Freshwater fishes of the USSR and adjacent countries, Part II. Izdatel'stvo Akademii Nauk SSSR, Moscow-Leningrad: 470–925. [In Russian]
- Bloch M.E. 1782. Oeconomische Naturgeschichte der Fische Deutschlands. 1. Theil, Berlin, 258 p.
- Bychowsky B. 1933. Notes about trematodes genus Dactylogyrus diesing crucian carps in ponds of park of Petergoff, Biological Research Institute. Trudy Biologicheskogo nauchno-issledovatel'skogo instituta LGU, 10: 269–296. [In Russian]
- Chang Y.-S., Huang F.-I. and Lo T.-B. 1994. The complete nucleotide sequence and gene organization of carp (*Cyprinus carpio*) mitochondrial genome. *Journal* of Molecular Evolution, 38: 138–155.
- Cherfas N.B. 1979. Gynogenesis in fish. In: V.S. Kirpichnikov (Ed.). Geneticheskie osnovy selektsii ryb [Genetic Bases of Fish Selection]. Nauka, Leningrad: 309–335. [In Russian]
- Chmilevsky D.A. and Ivanov A.A. 2005. Ichthyofauna of ponds. In: D.J. Vlasov (Ed.). Park "Sergievka" – kompleksnyi pamyatnik prirody [Sergievka Park is a Complex Nature Monument]. Saint Petersburg State University, Saint Petersburg: 131–133. [In Russian]
- Efron B. 1979. Bootstrap methods: Another look at the jackknife. *Annals of Statistics*, 7: 1–26.

- Grib A.V. 1935. Some information on the crucian carp from the pond of Petergof. *Trudy Petergofskogo Biologichesk*ogo Instituta, 13–14: 96–104. [In Russian]
- Hasegawa M., Kishino H. and Yano T. 1985. Dating of the human-ape splitting by a molecular clock of mitochondrial DNA. Journal of Molecular Evolution, 22: 160–174.
- Hensel K. 1971. Some notes on the systematic status of Carassius auratus gibelio (Bloch, 1782) with further record of this fish from the Danube River in Czechoslovakia. Věstník Českolovenské Společnosti Zoologické, 35: 186–198.
- Holčík J. and Žitňan R. 1977. On the expansion and origin of *Carassius auratus* in Czechoslovakia. *Folia Zoologica*, 27: 279–288.
- Hosoya K. 2002. Cyprinidae. In: T. Nakabo (Ed.) Fishes of Japan with Pictorial Keys to the Species, Tokai University Press, Tokyo: 253–271.
- Iguchi K., Yamamoto G., Matsubara N. and Nishida M. 2003. Morphological and genetic analysis of fish of a *Carassius* complex (Cyprinidae) in Lake Kasumigaura with reference to the taxonomic status of two all-female triploid morphs. *Biological Journal of the Linnean Soci*ety, **79**: 351–357.
- International Commission on Zoological Nomenclature (ICZN). 2003. Opinion 2027 (Case 3010): Usage of 17 specific names based on wild species which are pre-dated by or contemporary with those based on domestic animals (Lepidoptera, Osteichthyes, Mammalia): conserved. Bulletin of Zoological_Nomenclature, 60: 81–84.
- Kalous L., Šlechtová Jr. V., Bohlen J., Petrtýl M. and Švátora M. 2007. First European record of Carassius langsdorfii from the Elbe basin. Journal of Fish Biology, 70 (suppl. A): 132–138. doi: 0.1111/j.1095– 8649.2006.01290.x
- Kirillov F.N. 1956. Karasi yakutskie Carassius carassius jacuticus ssp. nova (Kirillov). In: F.E.Karantonis, F.N. Kirillov and F.B. Muhomedijarov (Eds.). Ryby srednego techeniya reki Leny [Fishes of the Middle Lena River] (Trudy Instituta Biologii Yakutskogo Filiala AN SSSR, 2): 90–98. [In Russian]
- Kottelat M. and Freyhof J. 2007. Handbook of European Freshwater Fishes. Kottelat and Freyhof, Cornol and Berlin, 646 p.
- Linnaeus C. 1758. Systema naturae per regna tria naturae, secundum classes, ordenes, genera, species, cum characteribus, differentiis, synonymis, locis. Tomus I. Editio decimal, reformata. Holmiae, 824 p.
- Murakami M., Matsuba C. and Fujitani H. 2001. The maternal origins of the triploid ginbuna (*Carassius auratus langsdorfi*): phylogenetic relationships within the *C. auratus* taxa by partial mitochondrial D-loop sequencing. *Genes and Genetic Systematics*, **76:** 25–32.
- Nakamura M. 1969. Cyprinid Fishes of Japan. Research Institute for Natural Resources, Tokyo, 455 p. [In Japanese]

- Paepke H.J. 1999. Bloch's Fish Collection in the Museum fur Naturkunde der Humboldt Universitat zu Berlin: an illustrated catalog and historical account. A. R. G. Gantner Verlag KG, Ruggel/Liechtenstein, 216 p., 32 pls.
- **Posada D. and Crandall K.A. 1998.** MODELTEST: testing the model of DNA substitution. *Bioinformatics*, **14**: 817–818.
- Saitou N. and Nei M. 1987. The neighbor-joining method: A new method for reconstructing phylogenetic trees. *Molecular Biology and Evolution*, 4: 406–425.
- Sakai H., Iguchi K., Yamazaki Y., Sideleva V.G. and Goto A. 2009. Morphological and mtDNA sequence studies on three crucian carps (*Carassius*: Cyprinidae) including a new stock from the Ob River system, Kazakhstan. *Journal of Fish Biology*, 74: 1756–1773.
- Swofford D.L. 2002. PAUP*: Phylogenetic analysis using parsimony (*and other methods), version 4.0b10. Sunderland: Sinauer Associates.
- Takada M., Tachihara K., Kon T., Yamamoto G., Iguchi K., Miya M. and Nishida M. 2010. Biogeography and evolution of the *Carassius auratus*-complex in East Asia. *BMC Evolutionary Biology*, **10**: 7.

- Thompson J.D., Higgins D.G. and Gibson T.J. 1994. CLUSTAL W: Improving the sensitivity of progressive multiple sequence alignment through sequence weighting position-specific gap penalties and weight matrix choice. Nucleic Acid Research, 22: 4673–4680.
- Vetešník L., Papoušek I., Halačka K., Lusková V. and Mendel J. 2007. Morphometric and genetic analysis of *Carassius auratus* complex from an artificial wetland in Morava River floodplain, Czech Republic. *Fisheries Science*, 73: 817–822.
- Vlasov D.J. (Ed.) 2005. Sergievka Park is Complex Nature Monument. Saint Petersburg State University, Saint-Petersburg, 143 p. [In Russian]
- Wu H.-W. 1977. Cyprinid Fishes of China, vol. 2. Shanghai People's Press, Shanghai: 598 + 109 p. [In Chinese]
- Yamamoto G., Takada M., Iguchi, K. and Nishida M. 2010. Genetic constitution and phylogenetic relationships of Japanese crucian carps (*Carassius*). *Ichthyological Research*, 57: 215–222.

Submitted June 17, 2011; accepted August 31, 2011.